Synthesis and Herbicidal Activities of Novel 3-(α-Hydroxymethylene) pyrrolidine-2,4-dione Derivatives Containing a Cyclopropane Moiety

You-quan Zhu,^{a*} Jin Zhang,^a Yan-wei Yuan,^a Li-fen Xie,^a Hai-zhen Xu,^b Xiao-mao Zou,^{a*} and Hua-zheng Yang^a

^aState Key Laboratory and Institute of Elemento-Organic Chemistry, Nankai University, Tianjin 300071, People's Republic of China ^bCollege of Chemistry, Tianjin Normal University, Tianjin 300387, People's Republic of China *E-mail: zyq8165@nankai.edu.cn

Received January 23, 2011

DOI 10.1002/jhet.909

Published online 20 March 2013 in Wiley Online Library (wileyonlinelibrary.com).



A variety of novel 3-(α -hydroxymethylene)pyrrolidine-2,4-dione derivatives containing a cyclopropane moiety were designed and synthesized in satisfactory yields. Their structures were confirmed by ¹H NMR and HRMS. The bioassays indicated that most of the title compounds displayed some extent herbicidal activities at 100 mg/mL.

J. Heterocyclic Chem., 50, 202 (2013).

INTRODUCTION

Heterocyclic compounds are significant in modern organic and agrochemical chemistry due to their presence in a majority of well-known pharmaceutical preparations [1]. For these reasons, we have for a long time been interested in the synthesis of new heterocycles of potential pharmaceutical value. Recently, our attentions were focused on the preparation of compound **1** through a modification of the natural product **2** [2–7], which was inhibitory toward 4-hydroxyphenylpyruvate dioxygenase (HPPD, EC 1.13.11.27) with an IC₅₀ = 18 μ M [8]. The bioassay results showed that when R^1 was electron-donating, compound **1** exhibited better herbicidal activities [3–7]. In particular, when R^1 , R^2 , and R^3 were 2,4-dimethoxy, *iso*-propyl, and hydrogen, respectively, compound **1** provided 93% control of *Echinochloa crus-galli* at pre-emergence at 187.5 g/ha.

It was also noticed that α -ketoisocaproate (**3**) serve as natural substrates for HPPD [9]. In light of these findings and subsequent comparisons of the molecular structures of both HPPD enzyme substrates and triketone type inhibitors, Lin and co-workers reported 2- cyclopropanoylcyclohexane-1,3-dione(**4**) exhibited higher *in vitro* inhibition activity against HPPD (IC₅₀ = 6 μ M) [10]. In order to find valuable new lead compounds with high herbicidal activities, compounds **5** were designed and synthesized through replacement of the aryl group of compound **1** by different cyclopropyl groups. In this paper, we described the synthesis and herbicidal activities of some 3-[(α -hydroxy(cyclopropyl)) methylene] pyrrolidine-2,4-diones **5** (Figure 1, Scheme 1).

RESULTS AND DISCUSSION

Preparations. Among the many methods for synthesizing β -keto esters of the type RCOCH₂CO₂C₂H₅, three classical

syntheses via acetoacetic esters [11], via Meldrum's acid [12,13], and via mixed malonic esters [14] are practically useful, although not always satisfactory in yield. In this paper, considering the properties of acyl chlorides (6) and the starting materials' availability, β -keto esters (8) were synthesized through a general and versatile method based on the noteworthy reactivity of Meldrum's acid.

Compound **5** can be synthesized by acetylation of pyrrolidine-2,4-dione followed by the aroyl group's migration [15] or reaction of aroyl acetates with *N*-alkyl aminoacetates [16]. We preferred the latter method to prepare the target products **5** for its less toxic reagents and greater convenience. Intermediate **8** was reacted with **9** in refluxing absolute xylene to give the target compound **5** (Scheme 1). This reaction was assumed to go through a nucleophilic addition/elimination reaction. During this process, an intermediate amide's formation and a cyclizative condensation were involved.

Structure–activity relationship. A series of the related target products **5** were prepared and their herbicidal activities were tested, and the results listed in Table 4 shows that most of the target products **5** exhibited a higher inhibition rate for *Brassica campestris* than that for *E. crus-galli* at $100 \,\mu$ g/mL and this result indicated the selectivity of this kind compounds was effected by the halogen's introduction(such as **5**s).

For *B.campestris* at 100 µg/mL, when the substituent R^3 was iso-propyl, propyl, allyl, and cyclo-propyl, the corresponding compounds **5** (except for **5m**, **5p**) generally exhibited better herbicidal activity than those compounds **5** (R^3 = butyl and sec-butyl) **2**), as a result of substituent steric hindrance.

For *E. crus-galli* at $100 \,\mu$ g/mL, most of the target compounds **5** exhibited moderate herbicidal activity, but



Figure 1. Chemical structures of compounds 1-5.



when R^1 , R^2 , and R^3 were methyl, methyl, and iso-propyl, respectively, compound **5s** showed about 89% inhibiting rate. Considering that compound **5s** also possessed excellent herbicidal activity (about 95%) for *B.campestris*, compound **5** (R^1 , R^2 = methyl) will be our next research emphasis.

In summary, a variety of novel $3-(\alpha-hydroxymethylene)$ pyrrolidine-2,4-dione derivatives were designed and synthesized in satisfactory yields and the bioassays indicated that most of the title compounds displayed some extent herbicidal activities.

EXPERIMENTAL

All chemicals were purchased from commercial sources and purified by recrystallization or distillation. NMR spectra were recorded on a Bruker DPX 400 spectrometer (Bruker BioSpin AG industriestrasse 26CH-8117, Fallanden, Switzerland.); data for ¹H are reported as follows: chemical shift (ppm), integration, multiplicity (s, singlet; d, doublet; t, triplet; q, quartet; m, multiplet; and br, broad), coupling constant (Hz), and number. The melting points were determined on an X-4 binocular microscope melting point apparatus (Beijing Tech Instruments Co., Beijing, China) and are uncorrected. High resolution mass spectra were recorded on a VARIAN 7.0 T direct inlet instrument (IonSpec Corporation, Lake Forest, USA). Solvents were fractionally distilled before use.

Compounds 8a-8d were synthesized as the literature described [12,13,17,18]. Meldrum's acid (0.01 M) was dissolved in 75 mL of methylene chloride, and the reaction mixture was placed under nitrogen and brought to 0°C with an ice bath; pyridine (0.02 M) was then added by means of a dropping funnel. The appropriate acid chloride (0.01 M) was added over a 30-min period to the vigorously stirred mixture. After the final addition, the solution was allowed to rise to room temperature and allowed to stand for 3 h. The reaction was monitored by TLC (silica gel). Upon termination, the mixture was then washed with 30 mL of 4 N HCl, 2% sodium bicarbonate, and distilled water saturated with sodium chloride. The dried (MgSO₄) organic phase was filtered and crude 7a-7d was obtained by solvent removal in vacuo and not further purified. The residue crude 7a-7d was resolved in 10 mL of ethanol and refluxed for 2 h. The solvent was removed, and the residue was purified by flash silica gel chromatography using petroleum ether (60-90°C) and ethyl acetate as the eluent to yield 8a-8d. All of the compounds were identified by ¹H NMR spectroscopy. Their yields of compounds **8a–8d** are listed in Table 1.

General synthetic procedure for 5a-5s. Compounds 5a-5s were synthesized as the literature described [6,7,16].

A mixture of 8 (3 mmol) and 9 (3.13 mmol) in dry xylene (5 mL) was heated at 125-130°C with stirring for 24 h. The cooled solution was added to methanolic CH₃ONa, prepared from Na metal (3.35 mmol) and methanol (4 mL) at room temperature with stirring. After the aforementioned mixture was stirred at room temperature for 48 h, water (20 mL) was added to the reaction mixture and the organic layer was separated and extracted twice with water. The original water layer and the extracts were combined and acidified to pH 2-3 with 2N HCl under cooling. The acidic solution was extracted three times with chloroform (30 mL), and the extracts were washed with saturated brine and then dried over Na2SO4. The solvent was removed under reduced pressure to give crude product 5, which was purified by flash column chromatography on silica gel, using ethyl acetate-petroleum ether as the eluant to afford the pure target product. The melting points and yields of compounds 5a-5s are listed in Table 2, and their ¹H NMR are listed in Table 3.

Bioassays. The herbicidal activities of the title compounds (**5a–5s**) were evaluated using a previously reported procedure [4,5].

Treatment. The emulsions of purified compounds were prepared by dissolving them in $100 \,\mu\text{L}$ of *N*,*N*-dimethylformamide with the addition of a little Tween 20 and proper water. There were two

Table 1

Yields and ¹H NMR data of compound 8. R^1 R^2 mp (°C) Yield (%) 8a Cl Cl 88 liquid 8b 82 Br Br liquid 1.21 (s, 3H), 1.28–1.31 (m, 6H), 2.12 (t, J=8.2 Hz, 1H), 2.25 (d, J=8.0 Hz, 1H), 3.54 (q, J=15.1 Hz, 2H), 4.21 (q, J=7.1 Hz, 2H),6.80 (d, J=8.5 Hz, 1H) CH₃ 8c CH₃ 44 liauid 8d CF_3 Cl 85 liquid 1.16 (s, 3H), 1.22 (t, J=7.1 Hz, 3H), 1.27 (s, 3H), 2.24 (t, J=8.5 Hz, 1H), 2.31 (d, J=8.0 Hz, 1H), 3.47 (q, J=14.7 Hz, 2H), 4.14 (q, J=7.1 Hz, 2H), 6.88 (d, J=9.1 Hz, 1H)

	Mething points, yields, and mass data of compounds 54 –55.					
	R^1	R^2	R^3	mp/°C	Yield (%)	MS (ESI, <i>m/z</i>):
5a	Cl	Cl	C_3H_7	liquid	58	330.0675[(M-H) ⁻ ,100%]
5b	Cl	Cl	iso-C ₃ H ₇	113-114	58	$330.0669[(M - H)^{-}, 100\%]$
5c	Cl	Cl	cyclo-C ₃ H ₅	liquid	18	$328.0515[(M - H)^{-}, 100\%]$
5d	Cl	Cl	allyl	liquid	58	$328.0513[(M - H)^{-}, 100\%]$
5e	Cl	Cl	C_4H_9	liquid	47	$344.0826[(M - H)^{-}, 100\%]$
5f	Cl	Cl	sec-C ₄ H ₉	liquid	56	$344.0826[(M - H)^{-}, 100\%]$
5g	Br	Br	C_3H_7	liquid	43	419.9639[(M-H) ⁻ ,100%]
5h	Br	Br	iso-C ₃ H ₇	liquid	51	443.9587[(M+Na) ⁺ ,100%]
5i	Br	Br	cyclo-C ₃ H ₅	liquid	27	$417.9485[(M - H)^{-}, 100\%]$
5j	Br	Br	allyl	liquid	50	$417.9482[(M - H)^{-}, 100\%]$
5k	Br	Br	C_4H_9	liquid	30	$433.9795[(M - H)^{-},100\%]$
51	Br	Br	sec-C ₄ H ₉	liquid	28	$435.9931[(M+H)^{-},100\%]$
5m	Cl	CF ₃	C_3H_7	54-55	60	364.0930[(M-H) ⁻ ,100%]
5n	Cl	CF ₃	iso-C ₃ H ₇	82-84	54	$388.0898[(M + Na)^+, 100\%]$
50	Cl	CF ₃	cyclo-C ₃ H ₅	liquid	37	$362.0772[(M - H)^{-}, 100\%]$
5p	Cl	CF ₃	allyl	liquid	41	$362.0771[(M - H)^{-}, 100\%]$
5q	Cl	CF_3	C_4H_9	liquid	32	$378.1089[(M - H)^{-}, 100\%]$
5r	Cl	CF ₃	sec-C ₄ H ₉	72-74	45	$378.1089[(M - H)^{-}, 100\%]$
5s	CH ₃	CH ₃	iso-C ₃ H ₇	liquid	54	290.1762[(M-H) ⁻ ,100%]

 Table 2

 Melting points yields and mass data of compounds 52–55

Table 3

¹H NMR of compounds **5a–5s**.

¹H NMR δ (ppm)

- **5a** 0.94 (t, *J* = 7.3 Hz, 3H), 1.35 (s,3H), 1.36(s,3H), 1.56–1.67(m,2H), 2.42(t, *J* = 8.8 Hz,1H), 3.23(d, *J* = 8.5 Hz, 1H), 3.40 (t, *J* = 7.2 Hz,2H), 3.72 (s, 2H), 6.23 (d, *J* = 9.3 Hz,1H)
- **5b** 1.14 (s, 3H), 1.15 (s, 3H), 1.28 (d, *J*=5.4 Hz, 6H), 2.34 (t, *J*=8.7 Hz, 1H), 3.15 (d, *J*=8.4 Hz, 1H), 3.61 (s, 2H), 4.38–4.45 (m, *J*=6.6 Hz, 1H), 6.16 (d, *J*=9.1 Hz, 1H)
- 5c 0.72–0.80 (m, 4H), 1.26 (s, 3H), 1.28 (s, 3H), 2.34 (t, *J*=9.0 Hz, 1H), 2.62–2.68 (m, 1H), 3.14 (d, *J*=8.4 Hz, 1H), 3.61 (s, 2H), 6.15 (d, *J*=9.1 Hz, 1H)
- **5d** 1.35 (s, 3H), 1.36 (s, 3H), 2.43 (t, *J*=8.8 Hz, 1H), 3.24 (d, *J*=8.4 Hz, 1H), 3.71 (s, 2H), 4.04–4.06 (m, 2H), 5.19–5.22 (m, 1H), 5.26–5.28 (m, 1H), 5.73–5.81 (m, 1H), 6.22 (d, *J*=9.2 Hz, 1H)
- **5e** 0.96 (t, *J* = 7.3 Hz, 3H), 1.31–1.40 (m, 2H), 1.35 (s, 3H), 1.36 (s, 3H), 1.53–1.60 (m, 2H), 2.42 (t, *J* = 8.7 Hz, 1H), 3.23 (d, *J* = 8.5 Hz, 1H), 3.44 (t, *J* = 7.2 Hz, 2H), 3.72 (s, 2H), 6.23 (d, *J* = 9.2 Hz, 1H)
- **5f** 0.82 (t, J = 7.4 Hz, 3H), 1.12 (d, J = 6.7 Hz, 3H), 1.25–1.29 (m, 6H), 1.42–1.51 (m, 2H), 2.34 (t, J = 8.8 Hz, 1H), 3.16 (d, J = 8.5 Hz, 1H), 3.49–3.62 (m, 2H), 4.11–4.20 (m, 1H), 6.15–6.19 (d-d, $J_1 = 4.8$, $J_2 = 9.2$, 1H)
- 5g 0.95 (t, *J*=7.4 Hz, 3H), 1.34 (s, 3H), 1.36 (s, 3H), 1.57–1.66 (m, *J*=7.3 Hz, 2H), 2.34 (t, *J*=8.7 Hz, 1H), 3.24 (d, *J*=8.4 Hz, 1H), 3.40 (t, *J*=7.3 Hz, 2H), 3.72 (s, 2H), 6.75 (d, *J*=8.9 Hz, 1H)
- **5h** 1.14 (s, 3H), 1.15 (s, 3H), 1.26–1.28 (m, 6H), 2.26 (t, *J*=8.6 Hz, 1H), 3.16 (d, *J*=8.4 Hz, 1H), 3.60(s, 2H), 4.37–4.46 (m, *J*=6.7 Hz, 1H), 6.68 (d, *J*=8.8 Hz, 1H)
- 5i 0.72–0.80 (m, 4H), 1.26 (s, 3H), 1.28 (s, 3H), 2.26 (t, J=8.6 Hz, 1H), 2.64–2.70 (m, 1H), 3.15 (d, J=8.5 Hz, 1H), 3.60 (s, 2H), 6.66 (d, J=8.8 Hz, 1H)
- **5j** 1.34 (s, 3H), 1.36 (s, 3H), 2.37 (t, *J*=8.6 Hz, 1H), 3.25 (d, *J*=8.4 Hz, 1H), 3.71 (s, 2H), 4.04–4.06(m, 2H), 5.19–5.22 (m, 1H), 5.26–5.29 (m, 1H), 5.72–5.81 (m, 1H), 6.74 (d, *J*=8.8 Hz, 1H)
- 5k 0.88 (t, *J* = 7.3 Hz, 3H), 1.24–1.33 (m, 8H), 1.46–1.53 (m, 2H), 2.27 (t, *J* = 8.6 Hz, 1H), 3.16 (d, *J* = 8.4 Hz, 1H), 3.37 (t, *J* = 7.1 Hz, 2H), 3.65 (s, 2H), 6.67 (d, *J* = 8.9 Hz, 1H)
- $\begin{array}{l} \textbf{51} \\ \textbf{0.83} (t, \textit{J}=7.4\,\textrm{Hz}, \textrm{3H}), 1.12 (d, \textit{J}=6.5\,\textrm{Hz}, \textrm{3H}), 1.27-1.29 (m, \textrm{6H}), 1.42-1.51 (m, \textrm{2H}), 2.26 (t, \textit{J}=8.6\,\textrm{Hz}, \textrm{1H}), 3.17 (d, \textit{J}=8.5\,\textrm{Hz}, \textrm{1H}), 3.50-3.64 (m, \textrm{2H}), 4.78-4.82 (m, \textrm{1H}), 6.69 (d, \textit{J}=5.3\,\textrm{Hz}, \textrm{1H}) \end{array}$
- **5m** 0.95 (t, J=7.3 Hz, 3H), 1.39 (s, 3H), 1.41 (s, 3H), 1.56–1.68 (m, J=7.2 Hz, 2H), 2.53 (t, J=9.0 Hz,1H), 3.36 (d, J=8.4 Hz, 1H), 3.41 (t, J=7.2 Hz, 2H), 3.74 (s, 2H), 6.89 (d, J=9.6 Hz, 1H)
- **5n** 1.14 (s, 3H), 1.15 (s, 3H), 1.31 (d, 6H), 2.44 (t, J=8.8 Hz, 1H), 3.27 (d, J=8.4 Hz, 1H), 3.62 (s, 2H), 4.38–4.44 (m, J=6.5 Hz, 1H), 6.81 (d, J=9.6 Hz, 1H)
- **50** 0.72–0.81 (m, 4H), 1.30 (s, 3H), 1.32 (s, 3H), 2.44 (t, J=9.0 Hz, 1H), 2.64–2.70 (m, 1H), 3.26 (d, J=8.4 Hz, 1H), 3.62 (s, 2H), 6.79 (d, J=9.6 Hz, 1H)
- **5p** 1.30 (s, 3H), 1.33 (s, 3H), 2.46 (t, *J*=9.0 Hz, 1H), 3.28 (d, *J*=8.42 Hz, 1H), 3.64 (s, 2H), 3.96–3.98 (m, 2H), 5.11–5.20 (m, 1H), 5.63–5.73 (m, 1H), 6.80 (d, *J*=9.6 Hz, 1H)
- **5q** 0.96 (t, *J* = 7.32 Hz, 3H), 1.29–1.41 (m, 8H), 1.53–1.60 (m, 2H), 2.52 (t, *J* = 9.0 Hz, 1H), 3.35 (d, *J* = 8.41 Hz, 1H), 3.45 (t, *J* = 7.40 Hz, 2H), 3.74 (s, 2H), 6.88 (d, *J* = 9.6 Hz, 1H)
- $5r \quad 0.83 \text{ (t, } J=7.4 \text{ Hz, } 3\text{H}\text{), } 1.12 \text{ (d, } J=6.8 \text{ Hz, } 3\text{H}\text{), } 1.27-1.29 \text{ (m, } 6\text{H}\text{), } 1.41-1.51 \text{ (m, } 2\text{H}\text{), } 2.44 \text{ (t, } J=8.9 \text{ Hz, } 1\text{H}\text{), } 3.28 \text{ (d, } J=8.5 \text{ Hz, } 1\text{H}\text{), } 3.51-3.63 \text{ (m, } 2\text{H}\text{), } 4.11-4.20 \text{ (m, } 1\text{H}\text{), } 6.80-6.84 \text{ (m, } 1\text{H}\text{)}$
- **5s** 1.19 (s, 3H), 1.21 (s, 3H), 1.30 (d, *J* = 18.3 Hz, 6H), 1.69 (s, 3H), 1.72 (s, 3H), 2.51 (t, *J* = 6.7 Hz, 1H), 2.76 (d, *J* = 5.6 Hz, 1H), 3.65 (s, 2H), 4.46–4.53 (m, *J* = 6.8 Hz, 1H), 5.02 (d, *J* = 7.9 Hz, 1H)

)

 Table 4

 Herbicidal activity of compounds 5a-5s (percent inhibition; concentration 100 me/mL)

	0 ,		
	Brassica campestris	Echinochloa crus-galli	
5a	94.8	30.0	
5b	94.8	44.3	
5c	92.2	42.0	
5d	94.8	44.3	
5e	75.9	41.4	
5f	87.0	40.0	
5g	74.0	35.0	
5h	83.7	22.0	
5i	79.8	15.0	
5j	92.8	74.1	
5k	60.3	12.8	
51	66.8	12.8	
5m	0	39.2	
5n	92.2	32.3	
50	85.7	43.1	
5p	0	35.0	
5q	5.7	33.1	
5r	60.3	23.7	
5s	94.8	89.4	

replicates for each treatment. The mixture of the same amount of water, *N*,*N*-dimethylformamide and Tween 20 was used as control.

Inhibition of the root growth of rape (B. campestris L). Rape seeds were soaked in distilled water for 4 h before being placed on a filter paper in a 6-cm Petri plate, to which 2 mL of inhibitor solution had been added in advance. Usually, 15 seeds were used on each plate. The plate was placed in a dark room and allowed to germinate for 65 h at 28 (\pm 1) °C. The lengths of 10 rape roots selected from each plate were measured, and the means were calculated. The percentage inhibition was used to describe the control efficiency of the compounds. The herbicidal activity is summarized in Table 4.

Inhibition of the seedling growth of barnyard grass (*E. crus-galli L. BeauV*). Ten *E. crus-galli* seeds were placed into a 50-mL cup covered with a layer of glass beads and a piece of filter paper at the bottom, to which 5 mL of inhibitor solution had been added in advance. The cup was placed in a bright room, and the seeds were allowed to germinate for 65 h at 28 (\pm 1) °C. The heights of the aboveground parts of the seedlings in each cup were measured and the means calculated. The percentage

inhibition was used to describe the control efficiency of the compounds. The herbicidal activity is summarized in Table 4.

Acknowledgments. We are grateful for financial support by the National Basic Research Program of China (No. 2010CB126103) and the National Natural Science Foundation of China (NO.20772066, NO.21072108).

REFERENCES AND NOTES

[1] Graupner, P. R.; Carr, A.; Clancy, E.; Gilbert, J.; Bailey, K. L.; Derby, J.-A.; Clifford, B. C. J Nat Prod 2003, 66, 1558.

 [2] Wu, Y. C.; Hu, F. Z.; Yang, H. Z. Chin J Pestic Sci 2001, 3(3), 1.
 [3] Zhu, Y. Q.; Si, X. K.; Zou, X. M.; Liu, B.; Yang, H. Z. Chin J Org Chem 2007, 27, 385.

[4] Zhu, Y. Q.; Zou, X. M.; Hu, F. Z.; Yao, C. S.; Liu, B.; Yang, H. Z. J Agric Food Chem 2005, 53, 9566.

[5] Zhu, Y. Q.; Liu, P.; Si, X. K.; Zou, X. M.; Liu, B.; Song, H. B.; Yang, H. Z. J Agric Food Chem 2006, 54, 7200.

[6] Zhu, Y. Q.; Yao, C. S.; Zou, X. M.; Hu, F. Z.; Liu, B.; Yang, H. Z. Molecules 2005, 10, 427.

[7] Zhu, Y. Q.; Hu, F. Z.; Zou, X. M.; Yao, C. S.; Liu, B.; Li, Y. H.; Yang, H. Z. Chin J Org Chem 2005, 25, 419.

[8] Meazza, G.; Scheffler, B. E.; Tellez, M. R.; Rimando, A. M.; Romagni, J. G.; Duke, S. O.; Nanayakkara, D.; Khan, I. A.; Abourashed, E. A.; Dayan, F. E. Phytochemistry 2002, 60, 281.

[9] Baldwin, J. E.; Crouch, N. P.; Lee, M.-H.; MacKinnon, C. H.; Zhang, Z. H. Bioorg Med Chem Lett 1996, 6, 1503.

[10] Lin, Y. L.; Wu, C. S.; Lin, S. W.; Yang, D. Y. Bioorg Med Chem Lett 2000, 10, 843.

[11] Shriner, R. L.; Schmidt, A. G.; Roll, L. J. 'Organic Syntheses', Collect. Vol. 2, Wiley, New York. N.Y., 1943, p 266; Guha, M.and Nasipuri, D. 'Organic Syntheses', Collect. Vol. 5, 1973, p 384; Viscontini M. and Merckling, N. Helv Chim Acta 1952, 35, 2280.

 [12] Yuji O.; Kiyoshi S.; Osamu Y. J Org Chem 1978, 43(10), 2087.
 [13] Frank D. M.; Giles D. M. Jr.; Richard T. B. J Agric Food Chem 1989, 37(2), 501.

[14] (a) Breslow, D. S.; Baumgarten, E.; Hauser, C. R. J Am Chem Soc, 1944, 66, 1286; (b) Taylor, E. C.; McKillop, A. Tetrahedron 1967,

23, 897; (c) Bowman, R. E.; Fordham, W. D. J Chem Soc, 1951, 2758;

(d) Pichat, L.; Beaucout, J.-P. Synthesis 1973, 537.

[15] Heather, J. B.; Milano, P. D. Acylated diketonic compounds. EP 0186117, 1986.

[16] Matsuo, K.; Kitaguchi, I.; Takata, Y.; Tanaka, K. Chem Pharm Bull 1980, 28, 2494.

[17] Jiang, H.; Zhang, J. M.; Du, W. Q.; Zhu, S. Z. Chin J Org Chem 2007, 25, 86.

[18] David D.; Sidney A. B. J Am Chem Soc 1948, 70, 3426.







Compound Details

Structure Search













Structure Search





5d

FC-1













5i

51









Structure Search















H₃C

5r













CH

H₃C





CH₃





9







Compound Details

Structure Search